

The *endo* and *exo* isomers of 1-(4-isopropyl-3,5-dioxa-8-aza- bicyclo[5.1.0]octan-8-yl)pyrrolidine- 2,5-dione

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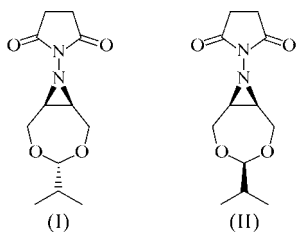
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The crystal structures of the title compounds, $C_{12}H_{18}N_2O_4$, have been determined in order to establish the relative configurations. In both structures, the isopropyl substituent strongly prefers the quasi-equatorial position. Therefore, the *exo* isomer adopts a dioxepanoaziridine pattern of a chair–chair (CC) conformation and the *endo* isomer a boat–chair (BC) conformation.

Comment

Fused aziridines, *e.g.* *N*-arenesulfonyl-protected 1,3-dioxepanoaziridines, are of interest as potent antihypoglycaemic agents (Dumić *et al.*, 1993, 1995; Oresić *et al.*, 2001; Filić *et al.*, 1996). Such compounds have been prepared from *trans*-acetylaminochlorodioxepanes by ring-closure dehydrohalogenation and subsequent *N*-protection with arenesulfonyl chlorides. Direct aziridination of 4,7-dihydro-1,3-dioxepines using sulfonyl azides as the nitrogen source failed (Dumić *et al.*, 1993). The crystal structures of these compounds have been thoroughly studied in order to develop quantitative structure–property and structure–activity relationship models.



Recently, we have developed a new aziridination procedure using *N*-aminosuccinimide as the nitrogen source and iodosylbenzene diacetate as the oxidizing agent, which directly leads to *N*-amino-protected fused dioxepanoaziridines (Flock *et al.*, 2005). Thus, a diastereomeric mixture of the title compounds, (I) and (II) (Figs. 1 and 2), was

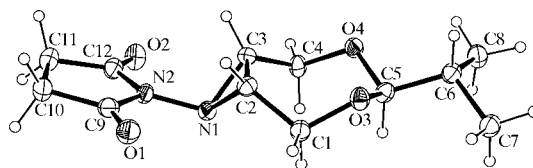


Figure 1

The *exo* isomer, (I), showing the atom-labelling scheme. Displacement ellipsoids are drawn at the 30% probability level and H atoms are shown as small circles of arbitrary radii.

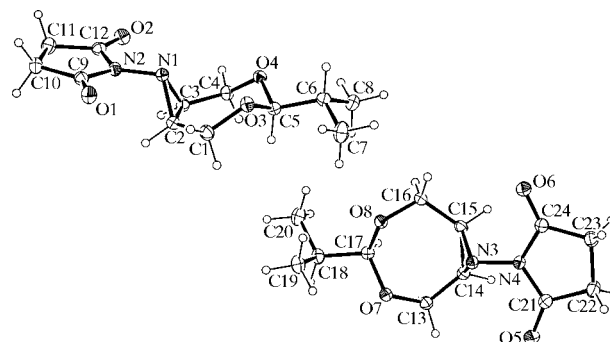


Figure 2

The asymmetric unit of the *endo* isomer, (II), showing the atom-labelling scheme. Displacement ellipsoids are drawn at the 30% probability level and H atoms are shown as small circles of arbitrary radii.

obtained from 2-isopropyl-4,7-dihydro-1,3-dioxepin, *N*-aminosuccinimide and iodosylbenzene diacetate. The relative configurations of these compounds are of interest with respect to their reactivity and stability.

Separation of the diastereomeric mixture by column chromatography afforded (I) and (II) as colourless solids. However, determination of the relative configurations using nuclear Overhauser experiments failed because of the flexibility of the dioxepinane moiety in solution. After recrystallization from diethyl ether–acetone and evaporation from acetone, respectively, (I) and (II) crystallized as well formed plate-like colourless crystals.

The *exo* isomer, (I), crystallizes in the non-centrosymmetric space group $P2_1$ with $Z' = 1$, but the absolute configuration could not be determined. The *endo* isomer, (II), crystallizes in space group $P2_1/c$ with $Z' = 2$. The difference between the two independent molecules in (II) lies in the conformation of the isopropyl substituent around the C5–C6 and C17–C18 bonds (Fig. 2). Neither isomer shows any strong intermolecular interactions.

In both isomers, the isopropyl substituent at the acetalic unit was found in a quasi-equatorial position. Analysis of the *endo* isomer, (II), reveals that the dioxepanoaziridine moiety adopts a boat–chair (BC) conformation, while for the *exo* isomer, (I), the preferred conformation in the solid state is the chair–chair (CC) conformation. The substituent on the aziridine N atom occupies, in both cases, the *anti* position with respect to the dioxepane ring, which supports previous results found for 8-sulfonyl-protected bicyclooctane derivatives (Dumić *et al.*, 1995).

Experimental

Under a nitrogen atmosphere, small portions of iodosyl benzene diacetate (3.865 g, 12 mmol) were added at room temperature to a solution of 2-isopropyl-4,7-dihydro-1,3-dioxepine (2.560 g, 18 mmol) and *N*-aminosuccinimide (1.141 g, 10 mmol) in dry acetonitrile (30 ml) over a period of 2.5 h. After stirring for a further 15 h, the reaction mixture was evaporated under reduced pressure. The oily residue was purified by column chromatography (silica gel, cyclohexane 100 → ethyl acetate–cyclohexane 80:20) to give the separated isomers, (I) and (II), as colourless solids (total yield 1.602 g, 6.3 mmol, 63%). Single crystals of isomer (I) (*exo*) for X-ray diffraction studies were obtained by slow evaporation from acetone (m.p. 413–415 K). ¹H NMR (500 MHz, CDCl₃): δ 0.90 [*d*, 6H, *J* = 6.84 Hz, CH(CH₃)₂], 1.84 [*m*, 1H, CH(CH₃)₂], 2.63 [*s*, 4H, CO–(CH₂)₂–CO], 2.82 [*m*, 2H, CH–N], 3.84 [*m*, 2H, O–CH₂], 4.12 [*d*, 1H, *J* = 6.84 Hz, O–CHR–O], 4.56 [*m*, 2H, O–CH₂]; ¹³C NMR (125 MHz, CDCl₃): δ 172.45 (2C, CO), 112.42 (1C, O–CHR–O), 66.64 (2C, O–CH₂), 47.46 (2C, CH–N), 31.56 [1C, CH(CH₃)₂], 26.46 [2C, CO–(CH₂)₂–CO], 17.74 [2C, CH(CH₃)₂]. Single crystals of isomer (II) (*endo*) for X-ray diffraction studies were obtained by recrystallization from diethyl ether–acetone (10:1 *v/v*) at 253 K (m.p. 415–417 K). ¹H NMR (500 MHz, CDCl₃): δ 0.88 [*d*, 6H, *J* = 6.88 Hz, CH(CH₃)₂], 1.83 [*m*, 1H, CH(CH₃)₂], 2.62 [*s*, 4H, CO–(CH₂)₂–CO], 2.71 [*m*, 2H, CH–N], 3.94 [*d*, 1H, *J* = 6.83 Hz, O–CHR–O], 3.98 [*m*, 2H, O–CH₂], 4.62 [*m*, 2H, O–CH₂]; ¹³C NMR (125 MHz, CDCl₃): δ 172.70 (2C, CO), 111.28 (1C, O–CHR–O), 65.36 (2C, O–CH₂), 48.15 (2C, CH–N), 32.23 [1C, CH(CH₃)₂], 26.60 [2C, CO–(CH₂)₂–CO], 17.72 [2C, CH(CH₃)₂]. APCI–MS, mixture of isomers: *m/z* (%) = 255 (100) [*M* + H]⁺; 287 (21) [*M* + H + MeOH]⁺.

Isomer (I)

Crystal data

C₁₂H₁₈N₂O₄ *D*_x = 1.363 Mg m⁻³
*M*_r = 254.28 Mo *K*α radiation
 Monoclinic, *P*2₁ Cell parameters from 7302 reflections
a = 6.1449 (11) Å *θ* = 1.6–26.0°
b = 7.8343 (10) Å *μ* = 0.10 mm⁻¹
c = 13.080 (3) Å *T* = 153 (2) K
β = 100.250 (15)° Plate, colourless
V = 619.6 (2) Å³ 0.47 × 0.41 × 0.05 mm
Z = 2

Data collection

Stoe IPDS-2 diffractometer 998 reflections with *I* > 2σ(*I*)
 Rotation scans *R*_{int} = 0.057
 Absorption correction: integration *θ*_{max} = 25.5°
 (*X-RED32*; Stoe & Cie, 2004) *h* = -7 → 7
*T*_{min} = 0.954, *T*_{max} = 0.995 *k* = -9 → 9
 8393 measured reflections *l* = -15 → 15
 1244 independent reflections

Refinement

Refinement on *F*² H-atom parameters constrained
R[*F*² > 2σ(*F*²)] = 0.036 *w* = 1/[σ²(*F*_o²) + (0.044*P*)²]
wR(*F*²) = 0.076 where *P* = (*F*_o² + 2*F*_c²)/3
S = 1.02 (Δ*σ*)_{max} = 0.004
 1244 reflections Δ*ρ*_{max} = 0.14 e Å⁻³
 165 parameters Δ*ρ*_{min} = -0.21 e Å⁻³

Isomer (II)

Crystal data

C₁₂H₁₈N₂O₄ *D*_x = 1.330 Mg m⁻³
*M*_r = 254.28 Mo *K*α radiation
 Monoclinic, *P*2₁/*c* Cell parameters from 10318 reflections
a = 24.470 (3) Å *θ* = 1.7–25.6°
b = 8.8444 (8) Å *μ* = 0.10 mm⁻¹
c = 11.8252 (14) Å *T* = 153 (2) K
β = 97.119 (9)° Plate, colourless
V = 2539.6 (5) Å³ 0.66 × 0.65 × 0.25 mm
Z = 8

Data collection

Stoe IPDS-2 diffractometer 2623 reflections with *I* > 2σ(*I*)
 Rotation scans *R*_{int} = 0.033
 Absorption correction: integration *θ*_{max} = 25.0°
 (*X-RED32*; Stoe & Cie, 2004) *h* = -29 → 29
*T*_{min} = 0.914, *T*_{max} = 0.972 *k* = -10 → 10
 18062 measured reflections *l* = -13 → 14
 4465 independent reflections

Refinement

Refinement on *F*² *w* = 1/[σ²(*F*_o²) + (0.0484*P*)²]
R[*F*² > 2σ(*F*²)] = 0.031 where *P* = (*F*_o² + 2*F*_c²)/3
wR(*F*²) = 0.084 (Δ*σ*)_{max} < 0.001
S = 0.90 Δ*ρ*_{max} = 0.21 e Å⁻³
 4465 reflections Δ*ρ*_{min} = -0.15 e Å⁻³
 330 parameters Extinction correction: *SHELXL97*
 H-atom parameters constrained (Sheldrick, 1997)
 Extinction coefficient: 0.0028 (4)

H atoms were treated as riding, with C–H distances in the range 0.98–1.00 Å and *U*_{iso}(H) = 1.2*U*_{eq}(C). The Friedel-equivalent reflections were merged.

For both compounds, data collection: *X-AREA* (Stoe & Cie, 2004); cell refinement: *X-AREA*; data reduction: *X-RED32* (Stoe & Cie, 2004); program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEP-3* (Farrugia, 1997); software used to prepare material for publication: *PLATON* (Spek, 2003).

Supplementary data for this paper are available from the IUCr electronic archives (Reference: GD1417). Services for accessing these data are described at the back of the journal.

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